

tion was characterized not by being the best at the early stages of the study, but rather by showing a very slow increase in R as the amount of data was increased. Further studies are planned as to the generality of this behavior.

The unusual features of the phytolaccagenin structure are its high degree of oxidation and the appearance of a carbomethoxy group. Other triterpene esters have been isolated,⁵ but their formation has been attributed to esterification during treatment with acidic alcohol. Since phytolaccagenin can be prepared without any exposure to methanol its ester function is apparently of natural origin.

Phytolaccagenin represents one of the largest natural products to be solved without the use of a heavy atom, and its solution exemplifies the growing possibilities for the use of chemical information in the early stages of an X-ray structural analysis.

Acknowledgments.—This work was supported in part by funds from the U. S. Public Health Service, Grant CA-04087, and by generous gifts of computer time from the Research Computer Laboratory, University of Washington. We also wish to thank Prof. C. E. Nordman for making available to us his model rotation program.

(5) C. Djerassi and J. S. Mills, *J. Am. Chem. Soc.*, **80**, 1236 (1958); W. A. Jacobs and O. Isler, *J. Biol. Chem.*, **119**, 155 (1937).

(6) National Science Foundation Cooperative Fellow, 1960-1961.

DEPARTMENT OF CHEMISTRY
UNIVERSITY OF WASHINGTON
SEATTLE 5, WASHINGTON

GEORGE H. STOUT
BERNARD M. MALOFSKY⁶
VIRGINIA F. STOUT

RECEIVED DECEMBER 5, 1963

Evidence that Cholesterol Sulfate is a Precursor of Steroid Hormones

Sir:

Since the initial observation¹ that conjugation of steroids does not necessarily destine the hormone for excretion, two important findings have supported the idea that intact steroid sulfates may serve as biosynthetic intermediates. In a recent study, Calvin, *et al.*,² found that urinary dehydroisoandrosterone sulfate (DS) may be synthesized *in vivo* from pregnenolone sulfate (PS) by a pathway involving intact steroid sulfates as intermediates. They have also shown³ that a homogenate of hyperplastic tissue is able to hydroxylate PS at C-17.

To extend these findings, cholesterol-7 α -³H sulfate-³⁵S (CS) (24.9×10^6 c.p.s. ³H and 4.35×10^6 c.p.m. ³⁵S, ³H-³⁵S ratio = 5.7) was injected into the left splenic artery of a female subject with an inoperable adrenal carcinoma. This vessel supplied 90% of the blood going to the neoplasm. The CS was injected through a small catheter introduced into the artery under radiographic visualization prior to the selective perfusion of the neoplasm with an antimetabolite.

The conjugates from the first 24-hr. urine collection were extracted into ethanol-ether (3:1).⁴ After evaporation of the organic solvents, the remaining aqueous phase was extracted with *n*-butyl alcohol. The dry residue left after removal of the butyl alcohol contained 2.5% of the injected radioactivity. It was purified by partition chromatography on Celite using the system⁵: isooctane-*t*-butyl alcohol-1 *M* NH₄OH (2:

(1) K. D. Roberts, R. L. VandeWiele, and S. Lieberman, *J. Biol. Chem.*, **236**, 2213 (1961).

(2) H. I. Calvin, R. L. VandeWiele, and S. Lieberman, *Biochem.*, **2**, 648 (1963).

(3) H. I. Calvin and S. Lieberman, *ibid.*, **3**, 259 (1964).

(4) R. W. J. Edwards, A. E. Kelly, and A. P. Wade, *Mem. Soc. Endocrinol.*, **2**, 53 (1952).

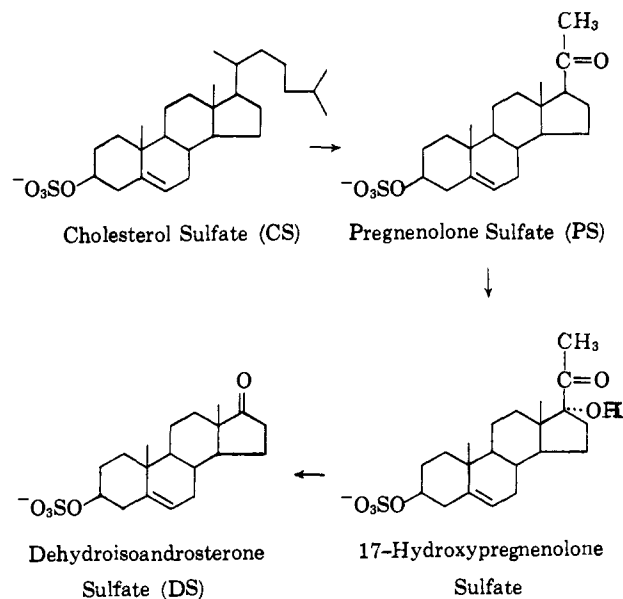
TABLE I
CRYSTALLIZATION DATA

Sample	Specific activity with resp. to		³ H- ³⁵ S ratio
	³ H in c.p.m./mg.	³⁵ S	
1st Crystallization	1134	212	5.4
Mother liquor	970	174	5.6
2nd Crystallization	1154	210	5.5
Mother liquor	1090	200	5.5
3rd Crystallization	1141	204	5.6
Mother liquor	1155	213	5.4

5:5), which separates the monosulfates from other urinary conjugates. The sulfate fraction was distributed between 0.3 *M* pyridinium sulfate solution and chloroform.⁶ Evaporation of the CHCl₃ afforded the pyridinium salts of the steroid sulfates (264,000 c.p.m. ³H and 40,000 c.p.m. ³⁵S) which were purified further by partition chromatography on Celite using the system: isooctane-chloroform-*n*-butyl alcohol-methanol-0.3 *M* pyridinium sulfate-pyridine (4:2:0.7:2:2:0.1). At least two radioactive metabolites were eluted, the more polar of which was rechromatographed on Celite using the system: isooctane-ethyl acetate-*n*-butyl alcohol-methanol-1 *M* NH₄OH (2:4:1:2:3). One crystalline radioactive product was eluted in the third to fifth hold-back volume, and this was identified by its infrared spectrum as the ammonium salt of DS. It weighed 87 mg. and contained 92,900 c.p.m. ³H and 17,100 ³⁵S. The product was recrystallized from a mixture of MeOH and acetone to constant specific activity with respect to both ³H and ³⁵S in both crystals and mother liquor residues during three successive crystallizations. The data are presented in Table I, where the ³H-³⁵S ratios are also shown. The latter values were essentially identical with the ratio present in the injected CS. In addition, the specific activities with respect to both isotopes remained constant following conversion of the DS to its oxime and two crystallizations of that derivative from methanol-ether. The yield of radioactivity associated with DS in the urine of the first day was approximately 0.5%.

SCHEME I

PROPOSED BIOSYNTHETIC PATHWAY INVOLVING STEROID SULFATES AS INTERMEDIATES



(5) P. K. Siiteri, Ph. D. Dissertation, Columbia University, 1963.

(6) J. McKenna and J. K. Norymberski, *Biochem. J.*, **76**, 1x (1960).

This demonstration of the conversion of CS to DS, a major secretory product of the adrenal,⁷ lends further support to the concept that sulfated intermediates, such as CS, PS, etc., are involved in the biosynthesis of the steroid hormones. Although the possibility of other pathways has not been excluded, the evidence available to date suggests that the conversion proceeds via C₂₁ intermediates as shown in Scheme I.

(7) R. L. VandeWiele, P. C. MacDonald, E. Gurrpide, and S. Lieberman, *Recent Progr. Hormone Res.*, **19**, 275 (1963).

(8) Predoctoral Research Fellow, National Institutes of Health.

(9) Career Scientist, Health Research Council, City of New York.

DEPARTMENTS OF BIOCHEMISTRY AND OBSTETRICS AND GYNECOLOGY
COLUMBIA UNIVERSITY
DEPARTMENT OF MEDICINE
NEW YORK UNIVERSITY
NEW YORK, NEW YORK

KENNETH D. ROBERTS
LAJOS BANDI
HAROLD I. CALVIN⁸
WILLIAM D. DRUCKER⁹
SEYMOUR LIEBERMAN

RECEIVED JANUARY 6, 1964

Identification of Peroxy Radicals by Electron Paramagnetic Resonance

Sir:

In e.p.r. studies free radicals are identified by their characteristic hyperfine splittings, their *g* values (line positions), and their relaxation times in a particular milieu. The last parameter is the least specific and the first parameter gives the most information. Peroxy radicals, RO₂·, have no hyperfine structure detectable by present methods except that due to O¹⁷. Because of the difficulty of O¹⁷ labeling only one experiment on the O¹⁷ hyperfine splittings of free radicals has been reported,¹ so that method is not yet broadly applicable.

We wish to report evidence that the *g* values of peroxy radicals easily distinguish them from RO· and R·. We have measured the *g* value of the cumyl peroxy radical which is 2.0155 ± 0.0005. Lontz² found that the radical CF₃CFCONH₂ reacts in the solid state with molecular oxygen to form a free radical without noticeable hyperfine splitting and with *g* values measured along the three principal axes of 2.0102, 2.0193, and 2.0267. He assumed this is the corresponding peroxy radical CF₃CFCONH₂. Fessenden and

O₂

Schuler³ found that alkanes irradiated with fast electrons in the absence of oxygen formed hydrogen atoms plus readily identifiable alkyl radicals. In the presence of oxygen under the same conditions radicals without noticeable hyperfine splitting were produced. The *g* values of these latter radicals ranged from 2.0155 when pentane was used to 2.0148 for paraffin oil, the *g* value decreasing with increasing molecular weight of the parent alkane. These were also presumed to be peroxy radicals.

Our cumyl peroxy radicals were produced in benzene solution as a result of the reaction of cumene with oxygen in the presence of azobisisobutyronitrile. No other radicals were detected in this system by e.p.r. Evidence that the principal radical formed in this system is in fact the cumyl peroxy radical, and the experimental arrangement, are discussed by Thomas.⁴

Ordinary free radicals of the type R· or RO·⁵ (or even R₂NO·¹) all have *g* values of less than 2.0100. Consequently, the location of the solution *g* values of peroxy radicals in the range 2.0140 to 2.0190 clearly

distinguishes them from these other radicals. One of McConnell and Robertson's generalizations⁶ originally applied to aromatic radicals, *i.e.*, the *g* value of a radical increases with the spin orbit coupling parameter, ζ , of the atoms that have significant spin density, applies also to this case.

To remove some spin density from the terminal oxygen atom and distribute it to any extent on the R group requires the polarization of the O-O bond so as to make the terminal oxygen more negatively charged. For this reason radicals with electron-withdrawing groups in R, such as the radical of Lontz, should have the greatest spin density on the terminal oxygen atom and therefore the highest *g* value. In solution this radical would have the averaged *g* value of 2.0187.

Our microwave frequency was measured by a Hewlett-Packard 524C counter with a Hewlett-Packard 540B transfer oscillator. The magnetic field was determined by proton magnetic resonance using a marginal oscillator whose frequency was measured on the same 524C counter. The line broadening effect of the presence of molecular oxygen was a principal source of error in the *g* value measurement.

(6) H. M. McConnell and R. E. Robertson, *J. Phys. Chem.*, **61**, 1018 (1957).

(7) University of Toronto, Toronto, Canada.

CALIFORNIA RESEARCH CORPORATION MALCOLM BERSOHN⁷
RICHMOND, CALIFORNIA J. R. THOMAS

RECEIVED JANUARY 8, 1964

Studies of Reactions of Atomic Carbon by a Double Tracer Technique

Sir:

Reaction mechanisms of atomic carbon have recently been studied using C¹¹ (20.5 min.) as produced by nuclear transformation.¹⁻⁴ The C¹¹, being radioactive, serves as its own tracer. However, it would also be greatly desirable to trace the origin of the atoms with which the C¹¹ combines. Because the nuclear recoil technique provides only a small number of carbon atoms (~10⁸) and hence of product molecules, this has not been practical thus far. Labeling the molecules with which the carbon reacts with another radioisotope is not feasible because the amount of such a secondary label which could be incorporated in the product is too small to detect.⁵

The use of stable isotopes, such as deuterium, as secondary tracers has faltered on the formidable problem of product analysis. Mass spectrometric separation of isotopic deuterated molecules followed by counting of their C¹¹ content is possible in principle, but very difficult in practice.⁶ The alternative, gas chromatographic separation of isotopic molecules followed by C¹¹ radioassay, has been restricted by the unavailability of columns of sufficient resolving power.

With the recent publication by Cvetanovic, *et al.*,⁷ of a technique for separation of deuterated light olefins

(1) C. MacKay, *et al.*, *J. Am. Chem. Soc.*, **84**, 308 (1962).

(2) C. MacKay and R. Wolfgang, *ibid.*, **83**, 2399 (1961).

(3) G. Stöcklin and A. P. Wolf, *ibid.*, **85**, 229 (1963).

(4) C. MacKay and R. Wolfgang, *Radiochim. Acta*, **1**, 42 (1962).

(5) In a typical experiment 10⁸ carbon atoms might react with a hydrocarbon labeled with tritium. Even if the hydrocarbon were completely labeled this would result in only an order of 10 to 100 tritium disintegrations per min. in the 10⁸ product molecules. Furthermore with a totally labeled molecule tritium self-radiolysis would be a serious problem.

(6) Mass spectrometric separation of trace isotopic product molecules would require high throughput efficiencies. Furthermore special counting techniques for the separated C¹¹ labeled molecules would have to be developed.

(7) R. J. Cvetanovic, F. J. Duncan, and W. E. Falconer, *Can. J. Chem.*, **41**, 2095 (1963).

(1) J. C. Baird, *J. Chem. Phys.*, **37**, 1879 (1962).

(2) R. Lontz, *Bull. Am. Phys. Soc.*, **8**, 328 (1963).

(3) R. W. Fessenden and R. H. Schuler, *J. Chem. Phys.*, **39**, 2147 (1963).

(4) J. R. Thomas, *J. Am. Chem. Soc.*, **85**, 591 (1963).

(5) M. S. Blois, H. W. Brown, and J. E. Maling in "Free Radicals in Biological Systems," Academic Press, New York, N. Y., 1961.